

Study of the hematological and some biochemical characteristics in common carp (*Cyprinus carpio* L.) fish feeding on fermented rations with varying concentrations growth stimulator (Bio boost aqua)

Al-Jubawi EY.A.¹; AL-humairi, K.O.M.^{1*}

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Abstract

The current study was conducted to find out the influence of yeast on fermented rations. Saccharomyces cerevisiae, and to which the growth enhancer Bio Boost Aqua was added in different proportions to take into account some hematological and biochemical characteristics for common carp fish. The hematological results of the studied fish before and at the end of the experiment showed a significant ($p \le 0.05$) superiority in T2 over the other treatments in RBC and PVC%, respectively (29.51±0.70; 2.85±0.07), while T1 and T2 recorded a significant decrease in leukocyte count at a rate of (25.70±0.14; 24.50±0.14) compared to the other experimental treatments before and at the end of the experiment. The results did not show any significant differences (p>0.05) in the hemoglobin HB levels of the test fish before and at the end of the experiment. The results of the analysis of white blood cells at the end of the experiment (neutrophils, eosinophils and monocytes) showed that there were no significant differences (p>0.05) in the experiments studied. While the white blood cells (the lymphocytes) in the control treatment showed higher rates with a rate of 36.01±1.45% compared to the other treatments which recorded lower values. The biochemical results in experimental fish before and at the end of the experiment showed an increasing value of total protein in blood serum in T2 and T3, which showed significant differences ($p \le 0.05$) at rates of (2.15, 1.95)%. Glucose levels showed a significant increase in T4 levels ($p \le 0.05$), reaching 76.5 mg/dL compared to the other experimental treatments where a decrease in levels was recorded before and at the end of the experiment. While the total cholesterol results for experimental fish before and at the end of the experiment showed that there were significant differences, T3 and T4 recorded higher values, reaching 76.49; 71.94 (mg/dl) compared to the other experimental treatments, which recorded a significant decrease in blood serum cholesterol analysis results $p \le 0.05$.

Keywords: Biochemical parameters, Total protein, Glucose, Total cholesterol

¹⁻Al-Furat AL. Awsat Technical University, AL- Musaib Technical College - Babylon, Iraq ^{*}Corresponding author's Email: kdm@atu.edu.iq

Introduction

The blood tests are one of the important metrics for assessing health and nutritional status and as an indicator of stress in fish culture (Nidhi and Meenakshi, 2022). The blood in fish is a red, liquid, sticky texture that enters the body through blood vessels. The percentage of blood volume is lower than that of the other vertebrates. Its size is 4 mil/100 g and this is due to the completion of the vascular system in teleost fish, which results in it requiring relatively less blood than the other vertebrates to carry out the vital processes in the body, it consists of plasma fluid, blood serum and Blood cells (Abdul Hamid, 2009). The study of blood indicators is of diagnostic importance. This includes examining the number of red blood cells, white blood cells, hemoglobin and packed cell size, all of which are influenced by environmental factors such as pollution from water pollutants. The blood indicators of fish are highly correlated with the fish's response to environmental and biological factors, which vary seasonally can affect changes in fish blood parameters such as reproductive cycle, nutrition, temperature, pH and photoperiod (Railo et al., 1985). It is also used as a diagnostic tool in biomonitoring, allowing us to detect physiological changes and pathogenesis resulting from diet, water quality, and disease (Adams et al., 1996). The blood tests are one of the indicators of fish physiological reactions. health. nutritional and environmental conditions (Iwashita et al., 2015). Nutritional status is a crucial factor in the host's defense against pathogens. The use of feed additives is therefore aimed not only at improving growth but also at improving the health and immune status of fish (Shokr and Mohamed. 2019). Fermentation is used to break down and digest the complex organic molecules to form smaller compounds and nutrients that are easily digestible (Nkhata et al., 2018) and this process can increase the digestibility of feed ingredients by analyzing the materials present in the feed ingredients through enzymes produced from microbes, fungi or yeasts through fermentation (Reisinger et al., 2012). Fermentation occurs through the use of baker's yeast S. cerevisieae in the feed and in turn improves growth and enhances the immunological response to the various fish (Tewary and Patra, 2011). Because it contains many immunostimulating compounds that are used as natural immunostimulants in the common nutritional systems of fish (Huyben et al., 2017). Use baker's yeast and natural growth enhancers to improve the growth performance of fish, and conversely, use some antibiotics and hormones as feed additives, which may cause harmful side effects to fish (Mahmoud et al., 2019). Repeated use of antibiotics in fish feed can also lead to the development of antibiotic-resistant bacteria and the formation of harmful substances that can pose a threat to the (Esiobu et al., 2002). ecosystem World Therefore, the Health Organization (WHO) has promoted the use of natural growth promoters such as yeasts and some herbs and medicinal plants to reduce the use of chemicals and replace them with natural sources. The use of natural materials is considered acceptable feed additives as they improve feed effectiveness and production performance (Levic et al., 2008). The use of baker's yeast S. cerevisieae in feed has been proven to improve growth and enhance the immune response of various fish species (Tewary and Patra, 2011). The research shows that baker's yeast has the ability to produce some energy substrates for intestinal cells, which makes the intestine healthy by improving the mucosal surfaces and microflora and improving the elongation of villi in the intestinal tract of fish (Lawrence et al., 2007).

Materials and methods

Cultural system. The current study used carp fish (Cyprinus carpio L.) with an initial weight between 36.03 and 36.5 grams, which came from a private farm in Babylon Governorate. The fish were then distributed at a rate of 10 fish per tank. with two replicates for each treatment, taking into account that the weights of the fish were as similar as possible in all experimental sessions. The experience lasted 60 days. The experiment was carried out in a recirculation system consisting of eight glass tanks (150 liters) with dimensions (100 x 40 x 50 cm). Fish were fed 3% of their body weight twice daily and fish weighing was performed every two weeks. During the experiment, some environmental factors of the water in the ponds were measured, such as: B.

Temperature (°C), dissolved oxygen concentration (mg/L), salinity (%) and pH value. Four diets were used in this experiment. The feed was ground well and homogeneously with an electric mill and then four feeds were formed, three of which were added with baker's yeast at an amount of 0.5 g/kg and added with water to form a dough. Thereafter, the three experimental diets were fermented for 48 hours using the method mentioned Radwan (2014). After by the fermentation of the three diets was completed, they were dried well, then two of them were taken out and ground well, and Bio Boost Aqua was added to them in the proportions shown in Tables 1 and 2, which represent the chemical composition of the diets used for feeding. Then it was put into a meat grinder to form pellets with a diameter of 2-3 mm, which corresponded to the size of the experimental fish's mouths. It was dried at room temperature for 72 hours, then stored in polyethylene bags with the name of the treatment and stored in the refrigerator at 4°C until use. The diets were formulated based on the treatments and distributed as follows:

- T1: treatment control diet without any treatment
- T2: fermented diet for 48 hours using S. cerevisiae at 0.5 g/kg
- T3: fermented diet and supplemented with 0.5 g/kg of the growth promoter (Bio Boost Aqua).
- T4: fermented feed and supplemented with 1 g/kg growth promoter (Bio Boost Aqua).

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| Componental/ | Experimental coefficients | | | | |
|---|---------------------------|------|------|-----------|--|
| Components% | T1 | Т2 | Т3 | T4 | |
| Vegetable protein concentrate (Carp-5 Extra | 10 | 10 | 10 | 10 | |
| Fish powder of Turkish origin | 5 | 5 | 5 | 5 | |
| Soybean meal (Argentinian(| 40 | 40 | 40 | 40 | |
| Flour | 14 | 14 | 14 | 14 | |
| yellow corn | 10 | 10 | 10 | 10 | |
| rice polishing | 10 | 10 | 10 | 10 | |
| Wheat Bran | 10 | 10 | 10 | 10 | |
| Premix | 1 | 1 | 1 | 1 | |
| S. cerevisiae | - | 0.05 | 0.05 | 0.05 | |
| Bio Boost Aqua | - | - | 0.05 | 0.1 | |

| Table 1: component of experimental diets and i | percentage (%) used in common carp |) fish feeding. |
|--|------------------------------------|-----------------|
|--|------------------------------------|-----------------|

 Table 2: the chemical composition % of the experimental diets according to the dry weight the values (Mean±SD) represent.

| Drovinato | Treatments | | | | |
|------------------|----------------------|------------------------|-------------------------|------------------------|--|
| composition (%) | First treatment | Second | Third | Fourth | |
| composition (70) | T1 | treatment T2 | treatment T3 | treatment T4 | |
| Moisture | 0.04 ± 6.26^{b} | 0.55±7.72 ^a | 0.04 ± 7.165^{a} | 0.06 ± 7.05^{a} | |
| Crude protein | 0.30 ± 28.88^{b} | 0.04 ± 31.87^{a} | 0.48±31.36 ^a | 0.09±31.11ª | |
| Fat | 1.11 ± 5.17^{a} | 0.61 ± 5.57^{a} | 0.35 ± 7.07^{a} | 0.39±6.55 ^a | |
| Ash | 0.29 ± 5.09^{ab} | 0.12±4.38 ^b | 0.22 ± 5.29^{ab} | 0.65 ± 5.98^{a} | |
| fiber | 0.40±12.91ª | 0.57 ± 7.36^{b} | 1.18 ± 5.96^{b} | 0.14 ± 6.78^{b} | |
| NFE | 1.57 ± 41.67^{a} | 0.67 ± 43.08^{a} | 0.88 ± 43.15^{a} | 0.24±42.51ª | |

*Different letters in the same row indicates significant differences ($p \le 0.05$) among the treatments in chemical content.

Hematological biochemical and analyzes. At the beginning of the experiment, blood samples were taken from the tail vein of various fish in the control group and at the end of the experiment, blood was taken directly from the heart of the experimental fish (3 fish/treatment) (cardiac puncture). with plastic syringes of size 2 mm. and do not use narcotic substances on the fish when collecting blood, as this could affect the accuracy of the blood tests on the test fish. This was done by collecting blood samples (1 ml from all fish) from the heart of each fish using a sterile syringe, previously flushed with ethylenediaminetetraacetic acid (EDTA) solution as an anticoagulant. as mentioned by (Opiyo et al., 2019). Place the collected blood samples into the tube and move it gently. Then close it with a twisting motion for (2-3) minutes and stick on it the number of treatments and repeat the samples, keep them cooled with ice and transfer them to carry out the analyses., including RBC, WBC, hemoglobin (Hb) and packed cell volume (PCV%), based on the mentioned methods (Coles, 1986). The second part of the blood samples collected from fish were collected without anticoagulant in Eppendorf tubes containing no anticoagulant (such as heparin) to obtain the blood serum. The tubes are then left in the cursive direction at room temperature for half an then ejected at 3000 hour and revolutions per minute for ten minutes, depending on the method (Yang and Chen, 2003). The serum was collected using a micropipette tube and then stored special tubes at -20°C. The in

biochemical tests were carried out using a spectrophotometer from Apel (Japan). by using private equipment from the company (Germany Human). The biochemical analyzes of the blood serum include the concentrations (total protein, glucose and total cholesterol).

Statistical Analysis: Statistical analysis was carried out using a completely randomized design (CRD) using the Statistical Analysis System (SAS, 2001) and the significant differences between the means of the characteristics were determined using Duncan's test (1955). Point tested level of (0.05).

Results

RBC and WBC number

The results of the current study demonstrate the importance of fermentation by baker's yeast *S. cereviseae* in the amount of 0.5 g/kg in the diet of common carp fish through its influence on blood indicators. This

appears in the results of the statistical analysis thereof, which showed in Table 3 the superiority of T2 ($p \le 0.05$) over all other treatments before and at the end of the experiment, in (RBC) with an average of 2.85×106 cell/mm³ and the clear superiority of T1, T3 and T4 in the results of the (RBC) analysis compared to the (RBC) results of the fish before the start of the experiment, which had 1.5 x 106 cells/mm³. and for (WBC), the results of statistical analysis confirmed significant differences ($p \le 0.05$) between the fish in T1 and T2 and other treatments before and at the end of the The (WBC) experiment. count decreased in T1 and T2 at a rate of 25.7×10³ 24.5×10 and cells/mm³. respectively. while no significant differences were found in T3 and T4 in the fish before the start of the experiment.

 Table 3: shows red blood cell count rates×10 ⁶ cells/mm³, white blood cell counts×10 ³ cells/mm³, and hemoglobin concentration (g/100 mL) and the percentage of the volume of packed blood cells of common carp fish fed on the experimental treatments for 60 days (Mean±SD).

| Hematological | Before the | Treatments at the end of the experiment | | | |
|---------------------------|----------------------|---|----------------------|-------------------------|------------------------|
| analyses | experiment | T1 (con.) | T2 | Т3 | T4 |
| RBC (10 ⁶ /µL) | 0.07 ± 1.55^{d} | 0.14 ± 2.50^{b} | 0.07 ± 2.85^{a} | $0.14 \pm 2.10^{\circ}$ | 0.07 ± 2.25^{bc} |
| WBC (10 ³ /µL) | 0.07 ± 26.45^{a} | 0.14±24.50° | 0.14 ± 25.70^{b} | 0.21 ± 26.45^{a} | 0.14 ± 26.61^{a} |
| Hemoglobin (g/dl) | 0.07 ± 8.35^{a} | 0.21 ± 8.45^{a} | 0.21 ± 9.15^{a} | 0.91 ± 6.65^{b} | 0.28 ± 6.80^{b} |
| PCV% | 0.70 ± 27.02^{b} | $0.70{\pm}27.50^{ab}$ | $0.70{\pm}29.51^{a}$ | 1.41±21.01° | $0.70\pm22.50^{\circ}$ |

*Different letters in the same row indicates significant differences ($p \le 0.05$) among treatments.

Hemoglobin concentration (Hb) and (PCV) %

The results of the analysis of hemoglobin concentration (Hb) and stacked cell size (PCV) in Table 3 showed significant differences ($p \le 0.05$) between the treatments before and at the end of the experiment. T1 and T2

of recorded the higher values hemoglobin concentration, reaching 8.45 and 9.15 g/100 ml, respectively, compared to the other treatments T3 and T4, which had lower values, which g/100 reached 6.65 and 6.8 ml, respectively. reached 100 ml. Concomitantly, there is also a significant

superiority of (PCV) in the same treatments, which achieved (27.5, 29.51) % in T1 and T2, respectively, compared to T3 and T4, which showed a decrease in (PCV) at average values of (21.01 and 22.50%, respectively.

White blood cells types

The types of white blood cells was showed in Table 4. The results of the statistical analysis showed that the number of white blood cells of lymphocyte type in T2, T3 and T4 recorded a significant decrease ($p \le 0.05$) compared to the control treatment T1, which recorded a higher value (36%). As for other types of white blood cells of the monocyte, eosinophil and neutrophil type, the results of the statistical analysis showed that there were no significant differences (p>0.05) between the levels of these cells in the control treatment and all other treatments examined.

Table 4: Explains the types of white blood cells in the blood of common carp fish in various fish.

| Turnes of (WDC) – | Experimental treatments | | | | |
|---------------------|-------------------------|-------------------------|--------------------------|--------------------------|--|
| Types of (WBC) | T1 (con.) | Τ2 | Т3 | T4 | |
| Lymphocyte | 36.01±1.45 ^a | 33.03 ± 1.414^{b} | 32.51±0.707 ^b | 33.53±0.707 ^b | |
| Monocyte | 3.01±0.705 ^a | 3.50±0.707 ^a | 3.02 ± 0.706^{a} | 3.50±0.707 ^a | |
| Eosinophil | 1.00 ± 0.703^{a} | 1.50 ± 0.707^{a} | 1.50 ± 0.707^{a} | 1.00±0.703 ^a | |
| Neutrophil | 60.00 ± 1.412^{a} | $62.00{\pm}1.414^{a}$ | 63.00 ± 1.414^{a} | 62.00 ± 1.414^{a} | |
| 1.001.14 | | | | | |

*Similar letters in the same row do not differ from each other significantly (p>0.05).

Serum total protein

The results of the serum total protein analysis for the experimental fish in Table 5 showed that there were significant differences ($p \le 0.05$) between T2, which recorded the highest value of 2.153 mg/dL, and the other experimental treatments before and at the end of the experiment, T3 followed at a rate of 1.954 grams/dl, while T1 and T4 showed no significant differences from the results of the analysis of the blood serum of the fish before the start of the experiment.

 Table 5: appears some Blood serum test standards For Common Carp fish that fed on different experimental diets (Mean± SD).

| At the beginning | Pland comm | The treatments at the end of experiment | | | |
|--------------------|-------------------------|---|--------------------------|--------------------------|--------------------------|
| Of experiment | blood serum | T1 | T2 | Т3 | T4 |
| Protein (g/dl) | 1.153±0.18° | 1.708 ± 0.0^{bc} | 2.153±0.073 ^a | 1.954±0.03 ^{ab} | $1.783 \pm 0.02^{\circ}$ |
| Glucose (mg/dl) | 46.19 ± 0.02^{b} | 49.45 ± 1.159^{b} | 55.32 ± 1.16^{b} | 54.095 ± 5.02^{b} | 76.50±2.31ª |
| Cholesterol(mg/dl) | $49.075 {\pm} 2.46^{b}$ | 51.55 ± 2.368^{b} | 49.525 ± 0.17^{b} | 76.49 ± 0.33^a | 71.94 ± 3.39^{a} |

*Characters differences in one row indicate a significant difference ($p \le 0.05$) among the treatments.

Glucose concentration

The results in Table 5 showed significant differences ($p \le 0.05$) in the glucose level of fish blood fed with feed containing the

growth promoter (BBA) at an amount of 1 gram/kg in T4. The glucose level increased in the fish of this treatment by 76.50 mg/dl compared to all other experimental treatments before and at the end of the experiment, in which no significant differences were found between each other (p>0.05) in blood sugar content in common carp fish.

Total cholesterol

The results of the analysis of total cholesterol in blood serum in carp fish shown in Table 5 showed a significant increase in cholesterol levels in T3 and T4, reaching 76.49 and 71.94 mg/dl, respectively, in comparison with the other treatments. It was also found that treatment T2 recorded a significant decrease ($p \le 0.05$) in cholesterol levels at a rate of 49.525 mg/dL compared to control treatment T1, which had 51.55 mg/dL, as well as the other treatments.

Discussion

The yeasts, through their enzymes that convert the proteins, have played an important role in converting the low nutrient biomass into higher value nutrient compounds at low cost (Couture et al., 2019; Lapena et al., 2020). in peptides and amino acids (Nkhata et al., 2018). Fermentation also has a better effect on the occurrence of biochemical changes in the food components, and the improvement of the food content of proteins and essential amino acids (Xiang et al., 2019) is promoted by the addition of baker's yeast S. cerevisiae to the food growth and enhances the immune response (Tewary and Patra, 2011). Some studies dealing with the evaluation of the nutritional value of baker's yeast S. cerevisiae in many fish species, including a study on (Korkmaz and Cakirogullari, 2011) on Nile tilapia fish and a study on Seabass fish by Peres et al., (1999) and also the study by (Tewary and Patra, 2011) on raw fish, all of these studies showed good results regarding the efficiency of using baker fish. s yeast in rations. The analyzes are carried out on blood contents to determine the health and physiological status of fish, which can serve as an indicator of disease or malnutrition and the stressful situations that occur due to undesirable nutritional or environmental factors (Witeska et al., 2016). Iwashita et al. (2015) pointed out that blood parameters of fish are indicators of health, well-being, physiological responses. nutritional and and environmental status. Blood tests are also considered valuable criteria for the nutritional situation as an indicator of stress in fish (Nidhi and Meenakshi, 2022). The analysis results of the blood tests carried out on the fish at the end of the experiment and the comparison with the blood tests before the start of the experiment showed a clear significant superiority of the fish fed with the T2 treatment compared to the other treatments. The results of the current study were consistent with the results of a study by Al-Refaiee et al. (2016) agree. This showed that adding yeast to fish rations increased the number of red blood cells in fish blood. This was confirmed by a study by Bransden et al. (2003) confirmed the addition of yeast in the diet of the Atlantic salmon fish Salmo salar L. can lead to an increase in the number of erythrocytes. While Witeska et al. (2015) pointed out that a

decrease in the number of red blood cells in carp fish beyond the normal limits of the species is evidence that fish are suffering from anemia. This in turn results in fish being unable to absorb large amounts of oxygen, even though it is present in the water. As a result, the fish suffer from a lack of oxygen (Ikun et al., 2013). Therefore, as a preventive measure, the fish resort to increasing the production of red blood cells in order to physiologically adapt to the environment and to support the continuous supply of the body tissues with oxygen present on the surfaces of the red blood is transported to cells (Mallya, 2007). White blood cells play the most important role in the defense mechanism of fish and are composed of monocytes, acid and neutrophils, lymphocytes and platelets (Nidhi and Meenakshi, 2022), the fish suffering from stress caused by environmental or nutritional changes increase the number of leukocytes (Hastuti and Subandiyono, 2015). The results of statistical analysis of the white blood cell count in the current study showed that there was a significant difference $(p \le 0.05)$ between the fish before the start and the end of the experiment, and a decrease in the white blood cell count was in the control treatment and T2 compared to the other experimental treatments before and at the end of the experiment observed that higher values were recorded in the number of white blood cells, this can possibly be attributed to the active role of the synergistic action of baker's yeast with growth promoter (Bio Boost) in T3 and T4 because it contains an immuneboosting formula. Opiyo et al. (2019), showed that blood samples from fed tilapia enriched with probiotics, brewer's yeast S. cerevisieae and Bacillus Spp. They contained a greater number of white blood cells than the control group. The results of the present study are consistent with the results of Hassaan et al. (2018) carried out study which involved the addition of β -glucan and nucleotide-rich yeast extract to Nile tilapia juvenile rations, treatments with this mixture were significantly better than the control diet. This benefit is due to the effect of the sugar's glucan, mannan and chitin on the cell wall of brewer's yeast, which help increase the number of white blood cells. Yeast is involved in the defense mechanism against pathogens, which has a positive effect on the immune parameters of the blood. Yeast also has an immunostimulant effect, fights diseases and increases the efficiency of phagocytosis and the effectiveness of the destruction of pathogenic bacteria and fungi (Schreck and Moyle, 1990). The results of the analysis of the concentration of hemoglobin Hb gm/dL and hematocrit % PCV showed a significant difference between the different experimental treatments before and after the experience. T1 and T2 recorded higher Hb gm/dL and PCV% values than T3 and T4, which recorded an observed decrease in Hb beyond the fish's natural limits. Research shows that the normal Hb concentration in teleost fish is between 7 and 10 g/100 ml (Abdul Hamid, 2009). In mixed treatments (T3, T4), the PCV% values decreased but

were within the acceptable range. Sources also indicate that the proportion of PCV in the blood of teleost fish varies between 20 and 40% (Hoar et al., 1992). This is due to the role of beneficial microorganisms that increase the absorption of iron, copper, folic acid, vitamins and other minerals (Van der Heuvel et al., 2000). White blood cells play a defensive role in the body against all types of pathogens, including fighting cancer cells. These cells are produced in the bone marrow and stored in the blood and lymphatic tissue, which contains various types of lymphocytes, monocytes eosinophils. and and neutrophils (Witeska et al., 2022). The results showed that there were no significant differences (p>0.05) between the white blood cell treatments tested: monocytes, eosinophils and neutrophils. Studies indicate that these cells are primarily responsible for the nonspecific immune response to infectious diseases in fish (Jeney and Anderson, 1993). The results of the research carried out clearly show that the use of brewer's yeast in experimental diets and the processing of the mixture, to which the growth stimulator Bio Boost was added in various proportions, did not affect the health of common carp fish. Research shows that when the proportion of these cells in fish exceeds the norm, this is considered an indicator of stimulation of the immune system against a particular disease. This was configured based on a study (Cengizler and Sahan, 2000). When fish are infected with diseases or parasites, a decrease in the number of red blood cells and an increase in the number

of leukocytes, including neutrophils are observed in infected fish. Lymphocytes come from blood cells, which are responsible for the production of antibodies in the bloodstream against any infection (Sahan and Duman, 2010). However, the decrease in the number of lymphoid cells in the experimental groups of this study indicates their activation and the absence of stimulation of fish Feeding with experimental treatments compared to the control treatment, clearly indicating the safe role of the fermentation process as well as the use of the natural growth stimulator

Bio Boost in feeding common carp. The study of serum parameters plays an important role in providing indicators of health, physiology and nutritional status, which help to determine the most feeding appropriate and breeding methods, making it possible to identify acute or chronic stress factors, as well as their manifestations to identify the disease (Peres et al., 1999). Whey proteins played an important role in the transfer of various nutritional materials and the defense of living beings against pathological factors, as well as in osmotic organization and some other tasks (Rudneva et al., 2011). Total protein results showed a significant benefit in T2 and T3, with rates higher than other experimental treatments where total protein decreased. This is due to the inability to synthesize proteins and limit the absorption or loss of proteins, thereby attenuating the effect in the blood (Patriche et al., 2011), and the exposure to stressful situations that occur in fish lead to increased energy

levels so that they can cope with environmental conditions to which toxic materials are exposed (Jenkins and Smith, 2003) The results of the present study are consistent with those of the study (Opiyo et al., 2019) in which the addition of brewer's yeast S. cerevisiae with a probiotic to Nile tilapia fish rations resulted in an increase of total protein content compared to the treatment control. However, the results of this study did not agree with the study results (Rozita et al., 2013). They reported that there were no significant differences in the concentration of total protein, albumin and globulin in serum after adding different amounts of brewer's yeast. on rationing Oscar fish (Astronotus ocellatus). This may be due to the active role of fermentation processes, which led to an increase in total protein concentration in the experimental diets. The sugar glucose is the main source of energy for most vertebrates and is a good indicator for checking the health of living beings (Jana et al., 2016). The results showed significant differences in glucose levels between the experimental groups. Compared to other experimental treatments, there was a significant increase in T4 before and after the end of the experiment. while there were no significant differences between them in the other treatments. Nemosok and Bores (1982) found that blood glucose appears to be a sensitive indicator of environmental and nutritional stress in fish. The results of the present study are consistent with the study by Abdel-Tawwab et al. (2008) agree. There is a significant increase in the percentage of glucose in the blood serum of tilapia fish fed with brewer's yeast compared to the control group, and the results of the study by Al-Refaiee et al. (2016), who demonstrated the superiority of brewer's yeast and growth stimulants, these results were attributed to the composition of the yeast cell wall, which contains chemical compounds, most of which are carbohydrates, beta-glucan and chitin, so the blood serum glucose The value increased after compliance of these diets around. The percentage of plasma cholesterol is associated with improved survival and health status of fish (Chatzifotis et al., 2004). The use of a growth promoter as a nutritional supplement to the feed ration inhibits pathogenic bacteria and also supports the immune response. Studies have shown increased cholesterol levels in fish fed a diet containing a mixture of probiotics and some growth promoters (Panigrahi et al., 2010; Piccolo et al., 2015). The results of the statistical analysis showed a significant increase in cholesterol levels in fish fed T3 and T4 compared to other experimental treatments before and after the end of the with significant experiment, no differences found between them. The results of the present study do not agree with the results of the study by Kowalska et al. (2015) agree. The results showed that the lowest blood serum cholesterol levels were found in the treatment in which yeast extract was added to 2% of feed ingredients and in another study in which iris trout were fed rations supplemented with 1-2 g/kg of yeast extracts. The results showed a significant reduction in total cholesterol compared to the control treatment (Mohebbi *et al.*, 2013).

Reference

- **Abdul Hamid, A.H.M., 2009.** Foundations of fish production and farming. College of Agriculture, Mansoura University, 1st edition, 640 P.
- Abdel-Tawwab, M., Abdel-Rahman, A.M. and Ismael, N.E.M., 2008. Evaluation of commercial live bakers' yeast, Saccharomyces cerevisiea as a growth and immunity promoter for fry Nile tilapia (Oreochromis niloticus L.) challenged in situ with Aeromonas hydrophila. Aquaculture, 280, 185-189.DOI:10.1016/j.aquaculture.2008.03. 055
- Admas, S., Ham, K.D., Greeley, M.S., Lehew, R.F., Hinton, D.E. and Saylor, CF., 1996. Downstream gradients in bio indicator responses point source contaminant effects on fish health. *Canadian Journal of Fisheries and Aquatic Sciences*, 53, 2177-2187. https://doi.org/10.1139/f96-191
- Al-Refaiee, I.H., Abdulrahman, N.M. and Mutter, H.A., 2016. Replacement of commercial dry yeast (*Saccharomyces cerevisiea*) with animal protein concentration and its effect in some blood parameters for fingerling common carp (*Cyprinus carpio*, L.). Basrah Journal of Veterinary Research, 15, 3,312-332.DOI: 10.23975 / bjvetr . 2016.172804.
- Bransden, M.P., Carter, C.G. and Nichols, P.D., 2003. Replacement of fish oil with sunflower oil in feeds for Atlantic salmon (*Salmo salar* L.). Effect on growth performance, tissue fatty acid composition and disease resistance. *Comparative Biochemistry & Physiolog* y - Part B, 135, 611–625. DOI:10.1016/s1096-4959(03)00143-x.

Cengizler, I. and Sahan, A., 2000. Living in Seyhan Dam Lake and Seyhan River determination of some blood parameters in mirror carps (*Cyprinus carpio*, Linnaeus, 1758). Turkish *Journal of Veterinary and Animal Sciences*, 24, pp. 205-214.

http://journals.tubitak.gov.tr/veterinary/ vol24/iss3/4

Chatzifotis, S., Muje, P., Pavlidis, M., Agren, J., Paalavuo, M. and Molsa, H., 2004. Evolution of tissue composition and serum metabolites during gonadal development in the common dentex (*Dentex dentex*). Aquaculture,236(1-4):557-573.

DOI:10.1016/j.aquaculture.2003.12.004

- **Coles, E.H., 1986.** Veterinary clinical pathology, 4thed.w.B. Sounders company Philadelphia , London, Toront, 43-64 .
- Couture, J.L., Geyer, R., Hansen,J.Ø., Kuczenski, B., Øverl, M. and Palazzo, J., 2019. Environmental benefits of novel nonhuman food inputs to salmon feeds. *Environmental Science & Technology*, 53, 1967-75. https://doi.org/10.1021/ acs.est.8b03832
- **Duncan, C.B., 1955.** Multiple range and Multiple, 'F' test. Biometric.,11:1-12.
- Esiobu, N., Armenta, L. and Ike, J., 2002. Antibiotic resistance in soil and water environments. *International Journal of Environmental Health Research*, 12(2),133-144.DOI:10.1080/ 09603120220129292
- Hassaan, M.S., Mahmoud, S.A., Jarmolowicz, S., El-Haroun, E.R., Mohammady, E.Y. and Davies, S.J., 2018. Effects of dietary baker's yeast extract on the growth, blood indices and histology of Nile tilapia (*Oreochromis niloticus* L.) fingerlings. *Aquaculture Nutrition*, 24(6), 1709-1717. DOI:10.1111/anu. 12805.
- Hastuti, S. and Subandiyono, J., 2015. Cat fish (*Clirias gariepnus* Barch) well-

being which maintained with bio floc Technolgy . *International Journal of Food Science & Technology*, 10(2), 74-90. http://www.bioflux.com.ro/aaci

- Hoar, W.S., Randall, D.J. and Farrell,A.P., 1992. Fish Physiology The Cardiovascular system. Vol. XII, Part. 474 P.
- Huyben, D., Vidakovic, A., Nyman, A., Langeland, M., Lundh, T. and Kiessling, A., 2017. Effects of dietary yeast inclusion and acute stress on postprandial whole blood profiles of dorsal aorta- cannulated rainbow trout. *Fish Physiology and Biochemistry*, 787-798. DOI:10.1007/s10695-016-0297-0.
- Ikun, F., Salosso, Y. and dan Toruan, L.N., 2013. Prosiding seminar National Kelautan dan Perikanan I (Universitas Nusa Cendana), 112-23.
- SAS., 2001. SAS User's Guide . Statistical . version . 6.12th ed . SAS. Institute . Inc . Cary. N.C . 7 P.USA.
- Iwashita, M.K.P., Nakandakare, I.B., Terhune, J.S., Wood, T. and Ranzani-Paiva, **M.J.T.** 2015. Dietary supplementation with Bacillus subtilis, Saccharomyces cerevisiae and Aspergillus oryzae enhance immunity and disease resistance against hydrophila Aeromonas and Streptococcus iniae infection in juvenile tilapia Oreochromis niloticus. Fish and Shellfish Immunology, 43, 60-66. DOI:10.1016/ j.fsi.2014.12.008.
- Jana, B., Pavel, H. and Libor, V., 2016. Glucose determination in fish plasma by two different moderate methods.*Acta Veterinaria Brunensis*, 85, 349–353 . DOI:10.2754/avb 201685040349.
- Jeney, G. and Anderson, D.P., 1993. Glucan injection of bath exposure given alone or in combination with a bacterin enhance the non-specific defence mechanisms in rainbow trout (*Oncorhynchus mykiss*). Aquaculture, 116(4), pp. 315-329.

https://doi.org/10.1016/0044-8486 (93)90416-V

- Jenkins, F. and Smith, J., 2003. Effect of sub lethal concentration of endosulfan on hematological and serum biochemical parameters in the carp, *Cyprinus carpio*. *Bulletin of Environmental Contamination and Toxicology*, 70, 993-947. DOI:10.1007/s 00128-003-0080-7
- Korkmaz, A.S. and Cakirogullari, G.C.,
 2011. Effects of partial replacement of fish meal by dried baker's yeast (*Saccharomyces cerevisiae*) on growth performance, feed utilization and digestibility in Koi carp (*Cyprinus carpio* L., 1758) fingerlings. *Journal* of *Animal and Veterinary* Adva nces, 10(3), 346-351.
- Kowalska, A., Zakęś, Z., Siwicki, A.K., Terech-Majewska, E., Jankowska, B., Jarmołowicz, S. and Głąbski, E., 2015. Impact of brewer's yeast extract and levamisole in diets with vegetable oils on the growth, chemical composition, and immunological and biochemical blood parameters of pikeperch (*Sander lucioperca*). *Czech Journal of Animal Science*, 60(11),498– 508.DOI:10.17221/8558 -CJAS
- Lapeña, D., Kosa, G., Hansen, L.D., Mydland, L.T., Passoth, V., Horn, S.J. and Eijsink, V.G.H., 2020. Production and characterization of yeasts grown on media composed of spruce - derived sugars and protein hydro lysates from chicken by-products. Microbial Cell Factories, 19: 1-14. 19. DOI:10.1186/s12934-020-1287-6
- Lawrence, P., Li, A., Castille, F.L. and Gatlin, D.M., 2007. Preliminary evaluation of a purified nucleotide mixture as dietary supplement for Pacific white shrimp (*Litopenaeus vannamei*). *Aquaculture Research*, 38, 887-890. DOI: 10.1111/j.1365-2109.2007.01761.x
- Lević, J., Siniša, M., Djuragić, O. and Slavica, S., 2008. Herbs and organic

acids as an alternative for antibioticgrowth-promoters. *Archivos de Zootecnia*, 11, 5-11.

- Mahmoud, R., Aziza, A., Marghani, B. and Eltavsh, R., 2019. Influence of ginger and garlic supplementation on performance, growth whole body composition and oxidative stress in the muscles of Nile tilapia (O. niloticus). Advances in Animal and Veterinary 397-404. Sciences, 75, http://dx.doi.org/10.17582/Journal.aavs/ 2019/7.5.397.404
- Mallya, Y.J., 2007. The effects of dissolved oxygen on fish growth in aquaculture. kingolwira national fish farming center, fisheries division. Ministry of Natural Resources and Tourism. Tanzania.,61 P.
- Mohebbi, A., Nematollahi, A., Gholamhoseini, A., Tahmasebi-Kohyani, A. and Keyvanshokooh, S., **2013**. Effects of dietary nucleotides on antioxidant status and serum the lipids of rainbow trout (Oncorhynchus mykiss). Aquaculture Nutrition, 19, 506 514 https://doi.org/10. 1111/anu.12002.
- Nemcsok, J. and Bores, L., 1982. Comparative studies on the sensitivity of different fish species to metal pollution. *Acta Biologica Hungarica*, 33, 23-27.
- Nidhi, S. and Meenakshi, S., 2022. Alteration in hematological and biochemical parameters in fresh water Catfish *Clarias batrachus* as compared to *Cyprinus carpio*. International Journal of Fisheries and Aquaculture Sciences, ISSN 2248-9975.Vol.12, Num.1, pp.1-14.
- Nkhata, S.G., Ayua, E., Kamau, E.H. and Shingiro, J.B., 2018. Fermentation And germination improve nutritional value of cereals and legumes through activation of endogenous enzymes . *Food Science & Nutrition*, 6, 2446– 2458. DOI:10.1002/ fsn3.846
- Opiyo, M.A., Jumbe, J., Ngugi, C. C., Charo-Karisa, H., 2019. Dietary

administration of probiotics modulates non - specific immunity and gut micro biota of Nile tilapia (*Oreochromis niloticus*) cultured in low input ponds. *International Journal of Veterinary Science and Medicine*, 7, 1, 1-9. DOI:10.1080/23144599.2019.1624299.

- Panigrahi, A., Kiron, V., Satoh, S. and Watanabe, T., 2010. Probiotic bacteria Lactobacillus rhamnosus influences the blood profile in rainbow trout Oncorhynchus mykiss (Walbaum). Fish Physiology and Biochemistr, 36, 969977. DOI:10.1007/s10695-009-9375-x
- Patriche, T.; Patriche, N.; Bocioc, E. and Coada, M. T., 2011. Serum

biochemical parameter of farmed carp (*C. carpio*). *International J. of the Bio flux Society*, 4(2):137-140

http://www.bioflux.com.ro/aaci

- Peres, H., Goncalves, P. and Oliva-Teles,
 A., 1999. Glucose tolerance in gilthead
 Sea bream (*Sparus aurata*) and
 European sea bass (*Dicentrarchus labrax*). Aquaculture, 179(1-4), 415–423. DOI:10.1016/S0044-486 (99) 00175-1
- Piccolo, G., Bovera, F., Lombardi, P., Mastellone, V., Nizza, S., Meo, C.D., Marono, S. and Nizz, A., 2015. Effect of *Lactobacillus plantarum* on growth performance and hematological traits of European sea bass (*Dicentrarchus labrax*). Aquaculture International, 23, 10251032. DOI:10.1007/S10499-014-9861-8
- Radwan, A.M. Abdel Moez, 2014. The effect of using yeast as growth stimulants in fish diets . Master thesis, College of Agriculture, Mashtohour. Benha University, department of Animal Production (Fish Nutrition), 123 P.
- Railo, E., Nikinmaa, M. and Soivio, A., 1985. Effects of sampling on blood parameters in the rainbow trout, Salmo gairdneri Richardson. Journal of Fish Biology, 26, 725-732.

https://doi.org/10.1111/j.1095-8649.1985.tb04312.x

- Reisinger, N., Ganner, A., Masching, S., Schatzmayr, G. and Applegate, T.J., 2012. Efficacy of a yeast derivative on broiler performance, intestinal morphology and blood profile. *Livestock Science*, 143, 195-200.DOI:10.1016/j.livsci.2011.09.013.
- Rozita, K., Seyed, M.M. and Mehran, J.B., 2013. Effects of *Saccharomyces cerevisiae* (Saccharomycetes : Saccharomycetaceae) on *Astronotus ocellatus* as growth promoter and immune stimulant. *International journal of the Bioflux Society*,6(6):587-598.
- Rudneva, I.I. and Kovyrshina, TB., 2011. Comparative study of characteristics electrophoretic of serum albumin of round goby Neogobius melanostomus from Black Sea and Azov Sea. International Journal of Advanced **Biological** Research. 2011;1(1):131-136.
- Sahan, A. and Duman, S., 2010. Effect of β Glucan on Hematology of Common Carp (*Cyprinus Carpio*) Infected by Ectoparasites. *Mediterranean Aquaculture Journal*, 1(1), 1-7. DOI:10.21608/maj.2010.2669.
- Schreck C.B. and Moyle P.B. (eds), 1990. Methods for Fish Biology. AmericanFisheriesSociety,Bethesda,US A.http://doi.org/10.47886/97809132355 84
- Shokr, E. and Mohamed, E., 2019. Effect of ginger on some hematological aspects and immune system in Nile Tilapia. *International*

Journal of Aquatic Biology, 12(1), 1-18.

- Tewary, A. and Patra, B.C., 2011. Oral administration of baker's yeast (*Saccharomyces cerevisiae*) acts as a growth promoter and immunomodulation in *Labeo rohita* (Ham.).*Journal* of *Aquaculture Researc h& Development*,2,1-7. DOI:10.4172/2155-9546.1000109
- Van der Heuvel, E.M., Schoterman, M.C. and Muijs, T., 2000. Transgalactooligosacchyides stimulate calcium absorption in postmenopausal women. *Journal of Nutria*,130,2938-2942. DOI:10.1093/jn/130.12.2938
- Witeska, M., Dudyk, J. and Jarkiewicz, N., 2015. Hematological effects of 2phenoxyethanol and intimidate in carp (*Cyprinus carpio* L.). Veterinary Anapestic Analogue, 42(5), 537-546.
- Witeska, M., Lugowska, K. and Kondera , E., 2016. Reference values of hematological parameters for Juvenile Cyprinus carpio. Bulletin of the European Association of Fish Pathologists, 36(4),169-180.
- Witeska, M., Kondera, E., Ługowska, K. and Bojarski, B., 2022. Hematological methods in fish–Not only for beginners. *Aquaculture*, 547, 17 P.
- Xiang, H.; Sun–Waterhouse, D., Waterhouse, G.I., Cui, C. and Ruan, Z., 2019. Fermentation enabled wellness foods: A fresh perspective. *Food Science and Human Wellness*, 8, 203–243. https://doi.org/10.1016/j.fshw.2019.08.0 03.
- Yang, J.L. and Chen, H., 2003. Serum metabolic enzyme activities and hepatocyte ultra-structure of common carp after Gallium exposure. *Zoology Studies*, 42(3), 455-461.